

pOG44

**Flp-recombinase expression vector designed for use
with the Flp-In™ System**

Catalog no. V6005-20

Version B
021402
25-0352



www.invitrogen.com
tech_service@invitrogen.com

Table of Contents

| | |
|------------------------------------|------------|
| Table of Contents | iii |
| Important Information | v |
| Methods | 1 |
| Overview | 1 |
| Using pOG44 | 3 |
| Appendix | 5 |
| pOG44 Vector | 5 |
| Technical Service | 7 |
| Purchaser Notification..... | 9 |
| References | 11 |

Important Information

Contents 20 µg pOG44, lyophilized in TE, pH 8.0

Shipping/Storage Lyophilized plasmid is shipped at room temperature and should be stored at -20°C.

Product Specifications The pOG44 vector is qualified by restriction enzyme digestion with specific restriction enzymes as listed below. Restriction digests must demonstrate the correct banding pattern when electrophoresed on an agarose gel (see below).

| Vector | Restriction Enzymes | Expected Results (bp) |
|--------|---------------------|-----------------------|
| pOG44 | <i>Kpn</i> I | 5438, 347 |
| | <i>Xba</i> I | 5785 |

Accessory Products Many of the reagents used in the Flp-In™ System are available separately from Invitrogen. See the table below for ordering information.

| Product | Amount | Catalog no. |
|----------------------|--------------------|-------------|
| pFRT/ <i>lacZeo</i> | 20 µg, lyophilized | V6015-20 |
| pFRT/ <i>lacZeo2</i> | 20 µg, lyophilized | V6022-20 |
| pcDNA5/FRT | 20 µg, lyophilized | V6010-20 |
| T7 Promoter Primer | 2 µg, lyophilized | N560-02 |
| Zeocin™ | 1 g | R250-01 |
| | 5 g | R250-05 |
| Hygromycin | 1 g | R220-05 |

Flp-In™ Expression Vectors Additional Flp-In™ expression vectors are available from Invitrogen. For more information about the features of each vector or to download a manual for a vector, refer to our World Wide Web site (www.invitrogen.com) or call Technical Service (see page 7).

| Product | Amount | Catalog no. |
|--|--------|-------------|
| pcDNA5/FRT/V5-His TOPO® TA Expression Kit | 1 kit | K6020-01 |
| pSecTag/FRT/V5-His TOPO® TA Expression Kit | 1 kit | K6025-01 |
| pEF5/FRT/V5 Directional TOPO® Expression Kit | 1 kit | K6035-01 |
| pEF5/FRT/V5-DEST Gateway™ Vector Pack | 6 µg | V6020-20 |

continued on next page

Important Information, continued

Flp-In™ Host Cell Lines

For your convenience, Invitrogen has available several mammalian Flp-In™ host cell lines that stably express the *lacZ-Zeocin*™ fusion gene from pFRT/*lacZeo* or pFRT/*lacZeo2*. Each cell line contains a single integrated FRT site as confirmed by Southern blot analysis. The cell lines should be maintained in medium containing Zeocin™. For more information about the Flp-In™ Cell Lines, see our World Wide Web site (www.invitrogen.com) or call Technical Service (see page 7).

| Cell Line | Amount | Catalog no. |
|----------------|-----------------------------------|-------------|
| Flp-In™-293 | 3 x 10 ⁶ cells, frozen | R750-07 |
| Flp-In™-CV-1 | 3 x 10 ⁶ cells, frozen | R752-07 |
| Flp-In™-CHO | 3 x 10 ⁶ cells, frozen | R758-07 |
| Flp-In™-BHK | 3 x 10 ⁶ cells, frozen | R760-07 |
| Flp-In™-3T3 | 3 x 10 ⁶ cells, frozen | R761-07 |
| Flp-In™-Jurkat | 3 x 10 ⁶ cells, frozen | R762-07 |

Methods

Overview

Introduction

pOG44 is a 5.8 kb Flp recombinase expression vector designed for use with the Flp-In™ System (Catalog nos. K6010-01 and K6010-02) available from Invitrogen. When cotransfected with the pcDNA5/FRT plasmid into a Flp-In™ mammalian host cell line, the Flp recombinase expressed from pOG44 mediates integration of the pcDNA5/FRT vector containing the gene of interest into the genome via Flp Recombination Target (FRT) sites. The vector contains the following elements:

- The human cytomegalovirus (CMV) immediate-early enhancer/promoter for high-level constitutive expression of the Flp recombinase in a wide range of mammalian cells (Andersson *et al.*, 1989; Boshart *et al.*, 1985; Nelson *et al.*, 1987)
- Synthetic intron to enhance expression of the *FLP* gene (Huang and Gorman, 1990; O'Gorman *et al.*, 1991)
- *FLP* gene encoding the Flp recombinase (Buchholz *et al.*, 1996) to mediate integration of the pcDNA5/FRT expression plasmid into the genome

For more information about the Flp-In™ System, the pcDNA5/FRT plasmid, and generation of the Flp-In™ host cell line, refer to the Flp-In™ System manual. The Flp-In™ System manual is supplied with the Flp-In™ Complete or Core Systems, but is also available for downloading from our Web site (www.invitrogen.com) or by contacting Technical Service (see page 7).

FLP Gene

The *FLP* gene was originally isolated from the *Saccharomyces cerevisiae* 2μ plasmid (Broach *et al.*, 1982; Broach and Hicks, 1980), and encodes a site-specific recombinase that is a member of the integrase family of recombinases (Argos *et al.*, 1986). The Flp recombinase mediates a site-specific recombination reaction between interacting DNA molecules via the pairing of interacting FRT sites. For more information about site-specific recombination, refer to the next page and published reviews (Craig, 1988; Sauer, 1994).

The native *FLP* gene encodes a protein of 423 amino acids with a calculated molecular weight of 49 kDa. The *FLP* gene expressed from pOG44 encodes a temperature-sensitive Flp recombinase which carries a point mutation (flp-F70L) that results in a change in amino acid 70 from phenylalanine to leucine (Buchholz *et al.*, 1996). For more information about the properties of the flp-F70L protein, see below and Buchholz *et al.*, 1996.

Activity of the Flp Recombinase

When tested in mammalian cells, the native Flp recombinase has been shown to possess optimum recombination activity near 30°C and relatively low activity at 37°C, a result consistent with its physiological role in yeast (Buchholz *et al.*, 1996).

The flp-F70L protein expressed from pOG44 exhibits increased thermostability at 37°C in mammalian cells when compared to the native Flp recombinase (Buchholz *et al.*, 1996). Studies have shown that the Flp recombinase expressed from pOG44 possesses only **10%** of the activity of the native Flp recombinase at 37°C (Buchholz *et al.*, 1996).

continued on next page

Overview, continued

Flp Recombinase-Mediated DNA Recombination

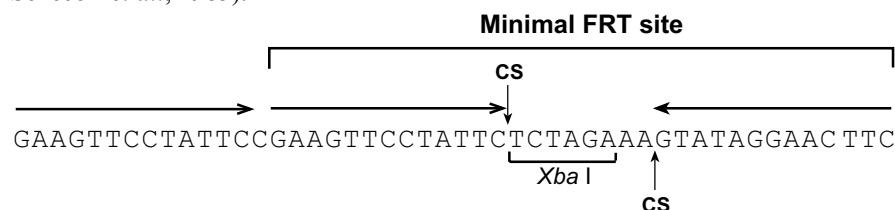
In the Flp-In™ System, integration of the pcDNA5/FRT expression construct containing your gene of interest into the genome occurs via Flp recombinase-mediated intermolecular DNA recombination. The hallmarks of Flp-mediated recombination are listed below.

- Recombination occurs between specific FRT sites (see below) on the interacting DNA molecules
- Recombination is conservative and requires no DNA synthesis; the FRT sites are preserved following recombination and there is minimal opportunity for introduction of mutations at the recombination site
- Strand exchange requires only the small 34 bp minimal FRT site (see below)

For more information about the Flp recombinase and conservative site-specific recombination, refer to published reviews (Craig, 1988; Sauer, 1994).

FRT Site

The FRT site, originally isolated from *Saccharomyces cerevisiae*, serves as a binding site for Flp recombinase and has been well-characterized (Gronostajski and Sadowski, 1985; Jayaram, 1985; Sauer, 1994; Senecoff *et al.*, 1985). The minimal FRT site consists of a 34 bp sequence containing two 13 bp imperfect inverted repeats separated by an 8 bp spacer that includes an *Xba* I restriction site (see figure below). An additional 13 bp repeat is found in most FRT sites, but is not required for cleavage (Andrews *et al.*, 1985). While Flp recombinase binds to all three of the 13 bp repeats, strand cleavage actually occurs at the boundaries of the 8 bp spacer region (see figure below) (Andrews *et al.*, 1985; Senecoff *et al.*, 1985).



CS = cleavage site

In the Flp-In™ System, the pFRT/*lacZeo* and pcDNA5/FRT vectors each contain a single FRT site. The pFRT/*lacZeo* plasmid is used to generate the Flp-In™ host cell line and the pcDNA5/FRT plasmid is used to express the gene of interest in the Flp-In™ host cell line. For more information about pFRT/*lacZeo*, pcDNA5/FRT, and the Flp-In™ System, refer to the Flp-In™ System manual.

Generating Stable Expression Cell Lines

You will cotransfect the pOG44 plasmid and your pcDNA5/FRT construct into your Flp-In™ host cell line(s) to generate stable cell lines that express your protein of interest. Cotransfection of pOG44 and pcDNA5/FRT allows expression of Flp recombinase resulting in integration of the pcDNA5/FRT plasmid into the genome via the FRT sites. Once the pcDNA5/FRT construct has integrated into the genome, the Flp recombinase is no longer required. The continued presence of Flp recombinase would actually be detrimental to the cells because it could mediate excision of the pcDNA5/FRT construct. For this reason, **the pOG44 plasmid lacks an antibiotic resistance marker for selection in mammalian cells.** When generating stable expression cell lines, the pOG44 plasmid and, therefore, Flp recombinase expression, will gradually be lost from transfected cells as they are cultured and selected.

Using pOG44

Introduction

General guidelines to transform pOG44 into *E. coli* are provided in this section.

General Molecular Biology Techniques

For help with *E. coli* transformation, restriction enzyme analysis, and DNA biochemistry, refer to *Molecular Cloning: A Laboratory Manual* (Sambrook *et al.*, 1989) or *Current Protocols in Molecular Biology* (Ausubel *et al.*, 1994).

E. coli Strain

Many *E. coli* strains are suitable for the propagation and maintenance of the pOG44 vector including TOP10, DH5 α , and JM109. We recommend that you propagate the vector in *E. coli* strains that are recombination deficient (*recA*) and endonuclease A deficient (*endA*).

For your convenience, TOP10 is available as chemically competent or electrocompetent cells from Invitrogen.

| Item | Quantity | Catalog no. |
|---|-----------------|-------------|
| One Shot [®] TOP10F' (chemically competent cells) | 21 x 50 μ l | C4040-03 |
| One Shot [®] TOP10 Electrocomp [™] (electrocompetent cells) | 21 x 50 μ l | C4040-52 |
| Electrocomp [™] TOP10 (electrocompetent cells) | 5 x 80 μ l | C664-55 |

Transformation Method

You may use any method of your choice for transformation. Chemical transformation is the most convenient method for many researchers. Electroporation is the most efficient and the method of choice for large plasmids.

Maintenance of Plasmid

To propagate and maintain the pOG44 vector, we recommend resuspending the vector in 20 μ l sterile water to prepare a 1 μ g/ μ l stock solution. Store the stock solution at -20°C.

Use this stock solution to transform a *recA*, *endA* *E. coli* strain like TOP10, DH5 α , JM109, or equivalent. Select transformants on LB agar plates containing 50 to 100 μ g/ml ampicillin. Be sure to prepare a glycerol stock of the plasmid for long-term storage (see below).

Preparing a Glycerol Stock

Once you have identified the correct clone, purify the colony and make a glycerol stock for long-term storage. You should keep a DNA stock of your plasmid at -20°C.

- Streak the original colony out on an LB plate containing 50 μ g/ml ampicillin. Incubate the plate at 37°C overnight.
 - Isolate a single colony and inoculate into 1-2 ml of LB containing 50 μ g/ml ampicillin.
 - Grow the culture to mid-log phase (OD₆₀₀ = 0.5-0.7).
 - Mix 0.85 ml of culture with 0.15 ml of sterile glycerol and transfer to a cryovial.
 - Store at -80°C.
-

continued on next page

Using pOG44, continued

Plasmid Preparation

Plasmid DNA for transfection into eukaryotic cells must be very clean and free from phenol and sodium chloride. Contaminants will kill the cells, and salt will interfere with lipid complexing, decreasing transfection efficiency. We recommend isolating plasmid DNA using the S.N.A.P.[™] MiniPrep Kit (10-15 µg DNA, Catalog no. K1900-01), the S.N.A.P.[™] MidiPrep Kit (10-200 µg DNA, Catalog no. K1910-01), or CsCl gradient centrifugation.



Several Flp-In[™] host cell lines which stably express the *lacZ-Zeocin*[™] fusion gene and contain a single integrated FRT site are available from Invitrogen (see page vi for ordering information). If you wish to express your gene of interest in 293, CV-1, CHO, 3T3, BHK, or Jurkat cells, you may want to use one of the Flp-In[™] host cell lines to establish your expression cell line.



We have observed down-regulation of the viral CMV promoter and subsequent loss of gene expression when pcDNA5/FRT-based expression constructs are introduced into 3T3 or BHK cells. This behavior is not observed with pEF5/FRT-based expression constructs. If you are generating Flp-In[™] expression cell lines using a 3T3 or BHK host cell line, we recommend that you clone your gene of interest into a pEF5/FRT-based expression plasmid (e.g. pEF5/FRT/V5-D-TOPO[®] or pEF5/FRT/V5-DEST). For more information, refer to our Web site (www.invitrogen.com) or call Technical Service (see page 7).



Because correct integration of your pcDNA5/FRT construct into the genome is dependent upon Flp recombinase, the expression levels of Flp recombinase in the cell will determine the efficiency of the recombination reaction. Flp recombinase levels must be sufficiently high to mediate recombination at the FRT sites (single recombination event) and overcome the low intrinsic activity of the enzyme (see page 1). We have varied the ratio of pOG44 and pcDNA5/FRT expression plasmid that we cotransfect into mammalian Flp-In[™] host cells to optimize the recombination efficiency. **We recommend that you cotransfect your Flp-In[™] host cell line with a ratio of at least 9:1 (w/w) pOG44:pcDNA5/FRT plasmid.** Note that this ratio may vary depending on the nature of the cell line. You may want to determine this ratio empirically for your cell line.



When transfecting your Flp-In[™] host cell line, be sure to use **supercoiled** pOG44 and pcDNA5/FRT plasmid DNA. Flp-mediated recombination between the FRT site on pcDNA5/FRT and the integrated FRT site in the Flp-In[™] host cell line will only occur if the pcDNA5/FRT plasmid is circularized. The pOG44 plasmid should be circularized to minimize the possibility of the plasmid integrating into the genome.

Cotransfection

Once you have cloned your gene of interest into pcDNA5/FRT and have prepared clean plasmid preparations of pOG44 and your pcDNA5/FRT construct, cotransfect the plasmids into your mammalian Flp-In[™] host cell line to generate your stable Flp-In[™] expression cell line. We recommend that you include the appropriate positive and negative controls to help you evaluate your results. Specific guidelines and protocols for generation of the Flp-In[™] expression cell line can be found in the Flp-In[™] System manual.

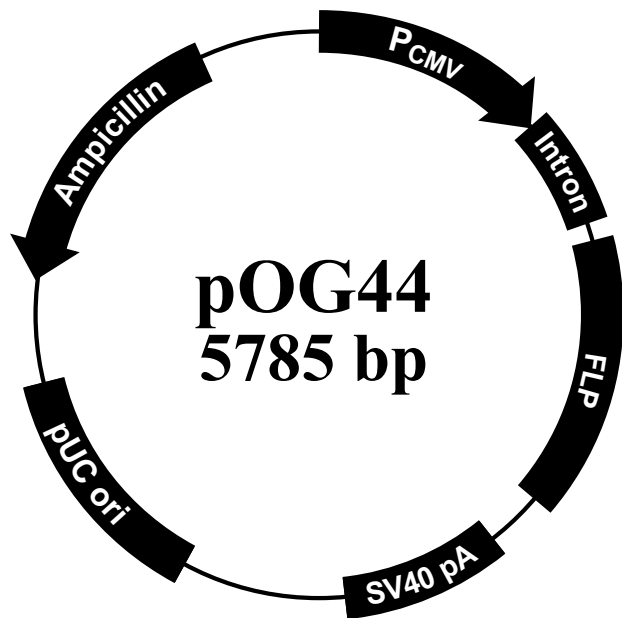
Reminder: The pcDNA5/FRT plasmid contains the hygromycin resistance gene to allow selection of transfectants using hygromycin. **The pOG44 plasmid does not contain an antibiotic resistance gene for selection in mammalian cells (see pages 5-6).**

Appendix

pOG44 Vector

Map of pOG44

pOG44 is a 5785 bp vector that expresses a temperature-sensitive Flp recombinase (flp-F70L) under the control of the human CMV promoter as previously described (O'Gorman *et al.*, 1991). The vector contains a synthetic intron to enhance expression of the *FLP* gene. Note that the vector does not contain an antibiotic resistance marker to allow stable selection in mammalian cells. The figure below summarizes the features of the pOG44 vector. **The complete sequence for pOG44 is available for downloading from our World Wide Web site (www.invitrogen.com) or by contacting Technical Service (see page 7).**



Comments for pOG44 5785 nucleotides

CMV promoter: bases 234-821

Synthetic intron: bases 871-1175

FLP ORF: bases 1202-2473

SV40 late polyadenylation signal: bases 2597-2732

pUC origin: bases 3327-3993 (complementary strand)

bla promoter: bases 4999-5097 (complementary strand)

Ampicillin (*bla*) resistance gene: bases 4138-4998 (complementary strand)

continued on next page

pOG44 Vector, continued

Features of pOG44 The table below describes the relevant features of pOG44. All features have been functionally tested.

| Feature | Benefit |
|---|--|
| Human cytomegalovirus (CMV) immediate early promoter | Allows high-level expression of the <i>FLP</i> gene (Andersson <i>et al.</i> , 1989; Boshart <i>et al.</i> , 1985; Nelson <i>et al.</i> , 1987) |
| Synthetic intron | Hybrid fragment which contains sequences derived from the adenovirus major late region and an IgG variable region (Huang and Gorman, 1990; O'Gorman <i>et al.</i> , 1991) and functions to enhance expression of the <i>FLP</i> gene |
| <i>FLP</i> ORF (flp-F70L) | Encodes a temperature-sensitive Flp recombinase (Buchholz <i>et al.</i> , 1996) that mediates conservative recombination via FRT sites (O'Gorman <i>et al.</i> , 1991) |
| SV40 late polyadenylation signal | Allows polyadenylation of mRNA |
| pUC origin | Allows high-copy number replication and growth in <i>E. coli</i> |
| <i>bla</i> promoter | Allows expression of the ampicillin (<i>bla</i>) resistance gene |
| Ampicillin (<i>bla</i>) resistance gene (β -lactamase) | Allows selection of transformants in <i>E. coli</i> |

Technical Service

World Wide Web



Visit the Invitrogen Web Resource using your World Wide Web browser. At the site, you can:

- Get the scoop on our hot new products and special product offers
- View and download vector maps and sequences
- Download manuals in Adobe® Acrobat® (PDF) format
- Explore our catalog with full color graphics
- Obtain citations for Invitrogen products
- Request catalog and product literature

Once connected to the Internet, launch your Web browser (Internet Explorer 5.0 or newer or Netscape 4.0 or newer), then enter the following location (or URL):

<http://www.invitrogen.com>

...and the program will connect directly. Click on underlined text or outlined graphics to explore. Don't forget to put a bookmark at our site for easy reference!

Contact Us

For more information or technical assistance, please call, write, fax, or email. Additional international offices are listed on our Web page (www.invitrogen.com).

United States Headquarters:

Invitrogen Corporation
1600 Faraday Avenue
Carlsbad, CA 92008 USA
Tel: 1 760 603 7200
Tel (Toll Free): 1 800 955 6288
Fax: 1 760 602 6500
E-mail:
tech_service@invitrogen.com

Japanese Headquarters

Invitrogen Japan K.K.
Nihonbashi Hama-Cho Park Bldg. 4F
2-35-4, Hama-Cho, Nihonbashi
Tel: 81 3 3663 7972
Fax: 81 3 3663 8242
E-mail: jpinfo@invitrogen.com

European Headquarters:

Invitrogen Ltd
3 Fountain Drive
Inchinnan Business Park
Paisley PA4 9RF, UK
Tel (Free Phone Orders): 0800 269 210
Tel (General Enquiries): 0800 5345 5345
Fax: +44 (0) 141 814 6287
E-mail: eurotech@invitrogen.com

MSDS Requests

To request an MSDS, please visit our Web site (www.invitrogen.com) and follow the instructions below.

1. On the home page, go to the left-hand column under 'Technical Resources' and select 'MSDS Requests'.
 2. Follow instructions on the page and fill out all the required fields.
 3. To request additional MSDSs, click the 'Add Another' button.
 4. All requests will be faxed unless another method is selected.
 5. When you are finished entering information, click the 'Submit' button. Your MSDS will be sent within 24 hours.
-

continued on next page

Technical Service, continued

Emergency Information

In the event of an emergency, customers of Invitrogen can call the 3E Company, 24 hours a day, 7 days a week for disposal or spill information. The 3E Company can also connect the customer with poison control or with the University of California at San Diego Medical Center doctors.

3E Company
Voice: 1-760-602-8700

Limited Warranty

Invitrogen is committed to providing our customers with high-quality goods and services. Our goal is to ensure that every customer is 100% satisfied with our products and our service. If you should have any questions or concerns about an Invitrogen product or service, please contact our Technical Service Representatives.

Invitrogen warrants that all of its products will perform according to the specifications stated on the certificate of analysis. The company will replace, free of charge, any product that does not meet those specifications. This warranty limits Invitrogen Corporation's liability only to the cost of the product. No warranty is granted for products beyond their listed expiration date. No warranty is applicable unless all product components are stored in accordance with instructions. Invitrogen reserves the right to select the method(s) used to analyze a product unless Invitrogen agrees to a specified method in writing prior to acceptance of the order.

Invitrogen makes every effort to ensure the accuracy of its publications, but realizes that the occasional typographical or other error is inevitable. Therefore Invitrogen makes no warranty of any kind regarding the contents of any publications or documentation. If you discover an error in any of our publications, please report it to our Technical Service Representatives.

Invitrogen assumes no responsibility or liability for any special, incidental, indirect or consequential loss or damage whatsoever. The above limited warranty is sole and exclusive. No other warranty is made, whether expressed or implied, including any warranty of merchantability or fitness for a particular purpose.

Purchaser Notification

Introduction

Use of the Flp-In™ System and its components (“System”) is covered under a number of different licenses including those detailed below.

The Nature of the Invitrogen License

Invitrogen Corporation (“Invitrogen”) has a license to sell the System to scientists **for research purposes only**, under the terms described below. Use of the System for any Commercial Purpose (as defined below) requires the user to obtain commercial licenses as detailed below. Note that each such license would cover only one part of the System. Before using the System, please read the terms and conditions set forth below. Your use of the System shall constitute acknowledgment and acceptance of these terms and conditions. If you do not wish to use the System pursuant to these terms and conditions, please contact Invitrogen’s Technical Services within 10 days to return the unused and unopened System for a full credit. Otherwise, please complete the User Registration Card and return it to Invitrogen.

Terms and Conditions

Invitrogen grants you a non-exclusive license to use the enclosed System for research purposes only. The System is being transferred to you in furtherance of, and reliance on, such license. You may not use the System, or the materials contained therein, for any Commercial Purpose without licenses for such purpose.

Definition of Commercial Purpose

Commercial Purpose includes:

- any use of the System or Expression Products in a Commercial Product;
- any use of the System or Expression Products in the manufacture of a Commercial Product;
- any sale of the System or Expression Products;
- any use of the System or Expression Products to facilitate or advance research or development of a Commercial Product; and
- any use of the System or Expression Products to facilitate or advance any research or development program the results of which will be applied to the development of a Commercial Product.

“Expression Products” means products expressed with the System, or with the use of any vectors or host strains in the System. “Commercial Product” means any product intended for sale or commercial use.

Individual Responsibilities

Access to the System must be limited solely to those officers, employees and students of your entity who need access to perform the aforementioned research. Each such officer, employee and student must be informed of these terms and conditions and agree, in writing, to be bound by same. You may not distribute the System or the vectors or host strains contained in it to others. You may not transfer modified, altered, or original material from the System to a third party without written notification to, and written approval from Invitrogen. You may not assign, sub-license, rent, lease or otherwise transfer any of the rights or obligations set forth herein, except as expressly permitted by Invitrogen.

continued on next page

Purchaser Notification, continued

Governing Law

This license shall be governed in its interpretation and enforcement by the laws of the United States and California.

FLP-Mediated Gene Modification

This product is licensed under U.S. Patent Nos. 5,654,182 and 5,677,177 and is **for research purposes only**. Inquiries about licensing for commercial or other uses should be directed to:

The Salk Institute for Biological Studies
10010 North Torrey Pines Road
La Jolla, CA 92037
Attn.: Department of Intellectual Property and Technology Transfer
Phone: 858-453-4100 ext 1275
Fax: 858-450-0509
Email: amueller@salk.edu

CMV Promoter

Use of the CMV promoter is covered under U.S. Patent Nos. 5,168,062 and 5,385,839 owned and licensed by the University of Iowa Research Foundation and may be used **for research purposes only**. Commercial users must obtain a license to these patents directly from the University of Iowa Research Foundation. Inquiries for commercial use should be directed to:

Brenda Akins
University of Iowa Research Foundation (UIRF)
214 Technology Innovation Center
Iowa City, IA 52242
Phone: 319-335-4549

References

- Andersson, S., Davis, D. L., Dahlbäck, H., Jörnvall, H., and Russell, D. W. (1989). Cloning, Structure, and Expression of the Mitochondrial Cytochrome P-450 Sterol 26-Hydroxylase, a Bile Acid Biosynthetic Enzyme. *J. Biol. Chem.* *264*, 8222-8229.
- Andrews, B. J., Proteau, G. A., Beatty, L. G., and Sadowski, P. D. (1985). The FLP Recombinase of the 2 Micron Circle DNA of Yeast: Interaction with its Target Sequences. *Cell* *40*, 795-803.
- Argos, P., Landy, A., Abremski, K., Egan, J. B., Ljungquist, E. H., Hoess, R. H., Kahn, M. L., Kalionis, B., Narayana, S. V. L., and Pierson, L. S. (1986). The Integrase Family of Site-Specific Recombinases: Regional Similarities and Global Diversity. *EMBO J.* *5*, 433-440.
- Ausubel, F. M., Brent, R., Kingston, R. E., Moore, D. D., Seidman, J. G., Smith, J. A., and Struhl, K. (1994). *Current Protocols in Molecular Biology* (New York: Greene Publishing Associates and Wiley-Interscience).
- Boshart, M., Weber, F., Jahn, G., Dorsch-Häsler, K., Fleckenstein, B., and Schaffner, W. (1985). A Very Strong Enhancer is Located Upstream of an Immediate Early Gene of Human Cytomegalovirus. *Cell* *41*, 521-530.
- Broach, J. R., Guarascio, V. R., and Jayaram, M. (1982). Recombination Within the Yeast Plasmid 2mu Circle is Site-specific. *Cell* *29*, 227-234.
- Broach, J. R., and Hicks, J. B. (1980). Replication and Recombination Functions Associated with the Yeast Plasmid, 2 mu Circle. *Cell* *21*, 501-508.
- Buchholz, F., Ringrose, L., Angrand, P. O., Rossi, F., and Stewart, A. F. (1996). Different Thermostabilities of FLP and Cre Recombinases: Implications for Applied Site-specific Recombination. *Nuc. Acids Res.* *24*, 4256-4262.
- Craig, N. L. (1988). The Mechanism of Conservative Site-Specific Recombination. *Ann. Rev. Genet.* *22*, 77-105.
- Gronostajski, R. M., and Sadowski, P. D. (1985). Determination of DNA Sequences Essential for FLP-mediated Recombination by a Novel Method. *J. Biol. Chem.* *260*, 12320-12327.
- Huang, M. T. F., and Gorman, C. M. (1990). Intervening Sequences Increase Efficiency of RNA 3' Processing and Accumulation of Cytoplasmic RNA. *Nuc. Acids Res.* *18*, 937-947.
- Jayaram, M. (1985). Two-micrometer Circle Site-specific Recombination: The Minimal Substrate and the Possible Role of Flanking Sequences. *Proc. Natl. Acad. Sci. USA* *82*, 5875-5879.
- Nelson, J. A., Reynolds-Kohler, C., and Smith, B. A. (1987). Negative and Positive Regulation by a Short Segment in the 5'-Flanking Region of the Human Cytomegalovirus Major Immediate-Early Gene. *Molec. Cell. Biol.* *7*, 4125-4129.
- O'Gorman, S., Fox, D. T., and Wahl, G. M. (1991). Recombinase-Mediated Gene Activation and Site-Specific Integration in Mammalian Cells. *Science* *251*, 1351-1355.
- Sambrook, J., Fritsch, E. F., and Maniatis, T. (1989). *Molecular Cloning: A Laboratory Manual*, Second Edition (Plainview, New York: Cold Spring Harbor Laboratory Press).
- Sauer, B. (1994). Site-Specific Recombination: Developments and Applications. *Curr. Opin. Biotechnol.* *5*, 521-527.
- Senecoff, J. F., Bruckner, R. C., and Cox, M. M. (1985). The FLP Recombinase of the Yeast 2-micron Plasmid: Characterization of its Recombination Site. *Proc. Natl. Acad. Sci. USA* *82*, 7270-7274.